$\ddot{}$

SYNTHESIS OF (\pm)-TALAROMYCINS A, B, C AND E

Raymond Baker*, Alastair L. Boyesf and Christopher 3. Swain*

***Chemistry Department, Merck Sharp and Dohme Research Laboratories, Neuroscience Research Centre, Terlings Park, Eastwick Road, Harlow, Essex, CM20 2QR +Department of Chemistry, The University, Southampton, SO9 5NH, U.K.**

<u>Summary</u> – Two key unsaturated spiroacetals have been used in the **synthesis of talaromycins A, B, C and E by routes involving chlorohydration and reductive dechlorination.**

The talaromycins A (1) and B (2) were isolated from the fungus Talaromyces stipitatus which grows on woodshavings-based animal feedstuffs. and were shown to be avian toxins11 Their activity arises through the blocking of outward potassium fluxes in smooth muscle leading to muscle dysfunction. In a more detailed re-examination of extracts from Talaromvces stipitatus, Lynn and co-workers isolated four more spiroketals, talaromycins C-F2 (3 - 5). talaromycins4 Both enantioselective3 and syntheses of the racemic have been reported. In this letter we report the stereodivergent synthesis of talaromycins A, B, C and E from comnon intermediates.

Talaromycin A (1)

Talaromycin B (2)

 $E(5)$ $F(6)$

985

Reagents: i) n-BuLi, THF, -78'C; ii) (8), 78%; iii) MeOH, Amberlyst-15, 100% iv) Ti2/Pd/CaCO\$Pb, MeOH; v) al-camphorsulphonic acid, CH2C12, 78%

Retrosynthetic analysis indicated the olefinic spiroacetal (7) to be an important intermediate. This spiroacetal could be prepared by the addition of a substituted δ -valerolactone (8) to the lithium anion of a suitably protected acetylenic diol (9), followed by partial hydrogenation of the keto-acetylene (10) to the Z-olefin and acid <code>catalysed spirocyclisation5</code> (Scheme 1). The acetylene (9) was prepared in 27% overall **yield from diethyl malonate in seven steps and was obtained as a colourless oil (bp 40-45Oc @ 2mniig)6.**

Addition of the lactone7 (g) to the lithium anion of acetylene (9) (Scheme 2) gave the hydroxyketo-acetylene (l0) in 79% yield as an oil (bp 117zC @ 0.06mmHg); 2220cm⁻¹ (C⊞C) and 1680cm⁻¹ (α,β-unsaturated carbonyl). Formation of the mixed methoxyacetal was accompanied by cleavage of the 1,3-dioxane ring to yield the acetylenic diol (l2) in 87% yield as a 3:l mixture of axial and aquatorial anomers which could be separated by flash chromatography on elution with EtOAc:petroleum (3:2+4:1) to afford the equatorial anomer as a colourless oil and the crystalline axial anomer (mp 73.5-74.5"C). Partial hydrogenation of the acetylenic diol (l2) and acid catalysed

SCHEME 3 Reagents: 1) t-BDMSCL, imidazole, *DMF*, σ_{ϕ} iii, the buocle, H₂, the side of the time of the time of the time iii) n-Bu₃SnH, AIBN (cat.), toluene; iv) n-Bu_ANF, THF

spirocyclisation gave the olefinic spiroacetals (*1*a,b) as a 2:1 mixture of inseparable **diastereomers (bp 62-64°C @ 0.04mmHg). The equatorial orientation of the ethyl group at** C-9 was assigned from the large coupling constants (10.8Hz) observed between H-8_{ax} and **H-Qax, for both (la) and (7b)). The H-8ax signal was a characteristic of subsequent** spiroacetals in this communication. Formation of the <u>tert</u>-butyldimethylsilyl ethers **(l3) and (l4) followed by careful flash chromatography (Et20:petrol; 1~19); enabled** each of the diastereomers (13) and (14) to be isolated in a combined yield of 84% (13) **Rf = 0..13 (bp 97-99°C @ 0.2mmHg),** $(\hat{14})$ **Rf = 0.18 (bp 97-99°C @ 0.2mmHg) (Scheme 3).**

The olefinic spiroketal (l3) was chlorohydrated to give the diastereomeric chlorohydrins (2) and (16) in a 1:4 ratio which were separated by flash chromatography *in* **73%** combined yield <u>[(15</u>) Rf = 0.28, Et₂0:petroleum, 1:4, H-4_{ax}, & = 4.19, ddd, J4a,OH = J4a,3e = 5Hz, J4a,5a = 10.8Hz; (<u>16</u>) Rf = 0.23, Et2D:petroleum, 1:4.
H-4_{eq}, δ = 3.78-3.72, m, including J_{4e,OH} = 5.2Hz]. Similarly, chlorohydration of **(l4) gave the m-diequatorial and w-diaxial chlorohydrins (17) and (l8) which** were obtained as colourless oils after flash chromatography (Et₂0:petroleum, 3:17, **(1_1) Rf = 0.21; (18) Rf = 0.39) in a ratio of 1.4:1 and a combined yield of 60%. The** large <u>trans</u>-diaxial coupling observed between H-3_{ax} and H-5_{ax} and H-4_{ax} in (17) **was particularly informative in assigning the relative stereochemistry at the three** contiguous centres C-3 to C-5 [(<u>17</u>), H-4_{ax}, & = 3.89, t, J_{4a.5a} = J_{4a.3a} = **9.9Hz; (B), H-4eq. 6 = 3.97, dt, J4e,DH = lO.BHZ, J4e,5e = J4e13a = 2.8Hz].** Both <u>trans</u>-diaxial chlorohydrins (<u>16</u>) and (<u>18</u>) exhibited sharp absorptions in their i.r.

of an spectra at _{vmax} 3480cm⁻' and 3510cm⁻' respectively, indicative of an
intramolecular hydrogen bond between the axial hydroxyl at C-4 and the O-7 in the **adjacent tetrahydropyran rfng, a phenomenon commonly observed in 4-hydroxy-1,7 dioxaspiro[5.5] undecanes8.**

Reductive de-chlorination9 of the individual chlorohydrins (15 - 18) gave the corresponding alcohols (19 - 22) as white crystalline solids mp's 56-583, 4%46.5'C, 68.5-70.5OC and 41-41.5V respectively and in yields of 60-80X. Finally, deprotection of the &-BDHS ethers (3 - 22) gave the talaromycins A, 6. C and E; all spectral data was in accordance with published data. Talaromycin B was isolated as a white crystalline solid mp 129.7-130.5°C C(CH2C12:hexane; 1:3), Lit4 127-128'C], whereas the other three talaromycins were isolated as colourless oils. This route to the talaromycins A, B, C and E & the olefinic spiroketals (l3) and (14) would be applicable to synthesis of the enantiomerically pure natural products by use of C-5 (R)-lactone (8).

Acknowledgements

We gratefully acknowledge financial support from the SERC and a CASE Award from Sigma Chemicals Company Ltd., (Poole).

References

- 1. **0.6. Lynn,** N.J. **Phillips, W.C. Hutton, J. Shabanowitz. I. Fennel1 and** R.J. **Cole, J-Am. Chem. Sec., 1982, 104, 7319.**
- **2. N.J. Phillips,** R.J. **Cole and D.G. Lynn, Tetrahedron Lett., 1987, 28, 1619.**
- **3. A.8. Smith** (III) **and A.S. Thompson, J. Org. Chem.. 1984, 49, 1469; M.H. Midland and J. Gabriel, J. Ora. Chem., 1985, 58, 1143; K. Mori and M. Kunaka, Tetrahedron, 1987, 43, 45; C. Iwata, M. Fujita, Y. Moritani, K. Hattori and T. Imanishi, Tetrahedron lett., 1987, 28, 3135;**
- **4. S.L. Schreiber and** T.J. **Sonnner, Tetrahedron Lett., 1983, 24. 4781; S.L. Schreiber,** T.J. Somer **and K. Satake, Tetrahedron Lett., 1985, 26, 17;** A.P. Kozikowski and J.G. Scripto, <u>J. Am. Chem. Soc</u>.,, 1984, 106, 353; P. Kocienski and C. Yeates, <u>J. Chem. Soc., Perkin Trans. I</u>, 1985, 1879; I.T. **Kay and D. Bartholomew, Tetrahedron Lett., 1984, 25, 2035;**
- 5. R. Baker, C.J. Swain and J.C. Head, <u>J. Chem. Soc. Chem. Commun</u>., 1985, 309.
- **6. Compound (ll) was idependently reported by M.A. Bates, J. Farina and M. Tong, 3_ Org. Chem.. 1986, 51, 2637.**
- **7. M.E. Kuehne, C.L. Kirkemo, T.H. Matsko and J-C. Bonhert, 3. Org. Chem., 1980, 45, 3259.**
- **8.** J.C. **Head, Ph.D. Thesis, University of Southampton, 1986.**
- **9. A similar approach was followed in the synthesis of milbemycin I33 by D.R. Williams, B.A. Barner, K. Nishitani and 3.6. Phillips, J. Am. Chem. Sot., 1982, 104, 4708.**

(Received in UK 30 December 1988)